

## MEASUREMENT OF SOIL WATERLOGGING TOLERANCE IN *PHASEOLUS VULGARIS* L.: A COMPARISON OF SCREENING TECHNIQUES

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Minnesota Agricultural Experiment Station Scientific Journal Series No. 12,111

(Accepted for publication 21 December 1982)

### ABSTRACT

Nelson, R.B., Davis, D.W., Palta, J.P. and Laing, D.R., 1983. Measurement of soil waterlogging tolerance in *Phaseolus vulgaris* L.: a comparison of screening techniques. *Scientia Hort.*, 20: 303-313.

Three methods not reported previously to determine flooding tolerance, and a scoring system developed for this study, were utilized to evaluate the effects of waterlogging upon 3 bean genotypes empirically differing in tolerance. These methods consisted of: (1) Triphenyl Tetrazolium Chloride Reduction Method (TTC), which measures flooding tolerance by determining the rate of root respiration during stress; (2) Electrical Conductivity Method (EC), in which the degree of root cell membrane injury is determined; and (3) Pressure Chamber Method (PC), by which the degree of desiccation is found by measuring the water potential of the xylem. A visual scoring method based on the external appearance of the plant was also used. The agreement among methods was generally good. The TTC and EC methods were more time-consuming and results more variable, especially from the latter. Results of the PC and scoring methods were highly correlated ( $r=0.85$ ). However, TTC and EC required 7-day-old seedlings, thus allowing the screening of large numbers of samples and efficient use of space. The  $F_1$  and  $F_2$  progeny from crosses between the most tolerant genotype (P074) and the most sensitive genotype (P149) indicated that differential reaction to flooding are probably heritable.

Keywords: *Phaseolus vulgaris* L.; soil waterlogging tolerance.

### ABBREVIATIONS

EC = Electrical Conductivity; PC = Pressure Chamber; TTC = Triphenyl Tetrazolium Chloride Reduction.

## RODUCTION

Common bean (*Phaseolus vulgaris* L.) production in some parts of the world is often limited by heavy rainfall, which frequently leads to waterlogged conditions in poorly drained soils (CIAT, 1977). During waterlogging, inhibition of root respiration and the subsequent energy transfer to the root system has been suggested as a cause of flooding stress (Rowe and Catlin, 1971). Alternatively, toxic effects of the products of anaerobic respiration have been suggested as the mechanism of root damage in both herbaceous and tree species (Crawford, 1966, 1967; Crawford and McManmon, 1968; Watts and Philipson, 1978).

Some of the effects of flooding resemble physiological drought (Kramer, 1961; Regehr et al., 1975; Broue et al., 1976). Thus, one consequence of flooding stress could be lowering of leaf water potential. A pressure chamber technique (Scholander et al., 1964) has been used intensively for rapid measurement of leaf water potential.

Loss of membrane integrity following flooding stress, resulting in leakage of solutes, has been reported by Grineva (1962). A conductivity method (Katter et al., 1932) measuring ion efflux following stress injury has been used successfully to evaluate relative freezing and heat-stress tolerance of several plant species (Li and Palta, 1978; Palta et al., 1981). Furthermore, ion leakage following stress injury has been reported to be due to specific alterations in the membrane transport system (Palta and Li, 1980).

Reduction of 2-3-5-triphenyl tetrazolium chloride, known as the TTC method (Kuhn and Jerchel, 1941; Kittock and Law, 1968), has been used to determine relative heat and freezing stress resistance (Steponkus and Lanar, 1967; Towill and Mazur, 1975; Palta et al., 1981).

The existence of genetic variability for flooding tolerance is well known (Kramer, 1961; Broue et al., 1976). Study of the genetic mechanisms associated with flooding tolerance has been the subject of several investigations (Kramer et al., 1977; Yoshida and Todano, 1978). Marshall and Millington (1977) inter-crossed 5 clover cultivars varying in sensitivity to flooded soils, and reported each cross to be more tolerant than either of the parents. Recently, Hely and Zorin (1977) reported an increase in waterlogging tolerance in alfalfa by subjecting plant material to rigid selection.

We investigated the potential of using TTC, conductivity and pressure-chamber techniques as rapid screening methods for measuring flooding tolerance.

## MATERIALS AND METHODS

*Plant material.* — Three bean (*Phaseolus vulgaris* L.) genotypes (P074, P254 and P149), differing in response to soil waterlogging, were chosen from among several lines that had been screened in preliminary studies following selection from the Centro Internacional Agricultura Tropical (CIAT).

From preliminary tests using visual scoring to determine tolerance, genotypes were classified as tolerant, sensitive and sensitive, respectively.

*TTC method.* — Twenty-five seeds of each genotype were germinated and grown inside a rolled moist paper towel. After 7 days, 3 seedlings from each genotype were selected for homogeneity of primary root length.

The terminal 5 mm of each root were discarded, and three 1-cm segments were excised and weighed immediately. Three root segments from one of the 3 seedlings within each line were then selected as control (non-stressed) samples and were immediately treated with TTC. The remaining 3 segments from each of 2 seedlings from each genotype were stressed to simulate flooded conditions by submersion in 10 ml of dd water and placed in a dark room at room temperature (22°C). Specially prepared wide-mesh screens were inserted over the segments to ensure complete submersion for the total length of the treatment. Three segments of one seedling from each genotype were thereafter removed successively at 12- and 24-h intervals following immersion and treated with TTC. The dd water used had about 6 mg l<sup>-1</sup> of dissolved oxygen, which was about 75% of the oxygen concentration of air. As root segments respire and use up the oxygen in water, the tissue experiences a progressively lower oxygen level.

The method for TTC treatment was similar to that of Palta et al. (1978). Briefly, the root segments were transferred at a desired time to test tubes containing 3 ml of 0.08% TTC in 0.05 M phosphate buffer of pH 7.4. The samples were incubated for 24 h in the dark at 22°C, the reduced TTC was extracted, and absorbance was recorded at 485 nm with a Bausch and Lomb 20 spectrophotometer. Data analysis was conducted according to a randomized complete-block design with 3 observations for each experimental unit and the 3 time-periods considered as blocks.

*EC method.* — The procedure used was similar to that of Palta et al. (1978). The root-segment samples were prepared as for the TTC test. Immediately after excision, all segments were individually transferred into 30-ml vials, submerged in 10 ml of dd water, and the electrical conductivity of the effusate in each vial recorded after the samples had been gently shaken for 1 h. This reading was taken as the non-stressed reference level. The vials thereafter remained undisturbed in a dark room at room temperature (22°C). After 8, 12, 24 and 36-h intervals following the initiation of flooding, conductance was read to determine the amount of ion leakage. After the final measurement, following 36 h of root submersion, the vials containing 10 ml of solution and root segment, were quickly frozen to about -20°C and were then thawed by transfer to tap water at about 25°C. This quick thaw procedure results in killing of the cells and releases all the ions into solution (Palta et al., 1981). The conductivity reading of the known tissue gave the total ion content of tissue. The amount of ion leakage due to flooding was determined by dividing the 8-, 12-, 24- and 36-h readings

reading of the killed tissue. Data were analyzed as from a randomized complete-block design.

*method.* — From each of the 3 genotypes, 96 uniformly-appearing seeds were planted in the greenhouse, 2 each in 12.5-cm diameter pots containing sterilized sand. Plants were thinned to one per pot and watered twice daily in a nutrient medium similar to Hoagland's solution (25 g  $\text{NH}_4\text{H}_2\text{PO}_4$ , 2 g  $\text{KNO}_3$ , 47.2 g  $\text{Ca}(\text{NO}_3)_2$ , 28.4 g  $\text{MgSO}_4$  and 0.34 g trace elements added to 132 l of tap water). Supplemental fluorescent lighting was supplied on a schedule of 16 h at an intensity of 60 mc, 6 dm above the plant tops. Temperature was maintained at 21/16°C day/night.

After 65 days, during which all of the flower buds of the plants had been continuously removed to allow more plant-to-plant developmental uniformity, 2 plants from each of the 3 genotypes were removed from the greenhouse and placed in a growth chamber with lighting and temperature conditions similar to that of the greenhouse.

The flooding treatment was produced by inserting the pots containing the plants into 16-cm diameter sealed plastic pots. Tap water was added to the outer container and maintained at a level 5 cm below the surface of the sand throughout the experiment. After 72 h, each plant was removed from the chamber, scored visually, and a pressure chamber was used to measure the stem water potential of the 3 largest leaves (Boyer, 1967). For this purpose, leaf sampling and preparation consisted of excising the most recently expanded leaf and placing it in a small plastic bag to reduce loss of water during measurement. This experiment was repeated 4 times. Each time, 2 plants were taken from each genotype.

In another experiment, the relationship between visual appearance and xylem water potential of 5 plants from each genotype was determined during 72 h of stress. The visual evaluation of such external symptoms as wilting and plant desiccation, discoloration, curling and collapse was accompanied by establishing a 1–6 stress rating. A rating of "1" indicated no symptoms, and each whole number thereafter represented a 20% increase. These data were then correlated on an individual plant basis with the xylem water potential measurement.

*Heritance studies.* — The 3 genotypes were used as parents in preliminary heritance studies.  $F_1$ ,  $F_2$  and back-cross populations were generated and raised in the greenhouse. Cultural practice and environmental control were similar to that of the pressure-chamber experiment described above. A randomized block design with 4 replications was used. Each block contained three 11.5 cm-diameter pots, representing each of the parental,  $F_1$  and reciprocal back-cross generations, and 8 pots of each of the 4  $F_2$  generations, i.e. including reciprocals.

The flooding treatment was applied in a similar way to the procedure described under pressure-chamber experiment. The data from each generation

within the 4 populations were tabulated to observe progeny distribution and segregation. The presence of significant phenotypic deviations from mid-parental value of the genotypes evaluated was tested by using the Least Significant Difference.

## RESULTS AND DISCUSSION

*TTC method.* — The TTC method was effective ( $P \leq 0.05$ ) in differentiating among the 3 genotypes, although no effect was perceived among the periods of stress (Fig. 1). P074 demonstrated a very high TTC reducing ability, while P524 demonstrated a rapid decline during submersion for 24 h; P149 was intermediate (Fig. 1). There was uniformity ( $P > 0.05$ ) among the 3 samples within the same genotype and stress period. Except for P149, the differences in tolerance among the genotypes, based on the TTC method, agreed with visual rating (Table I).

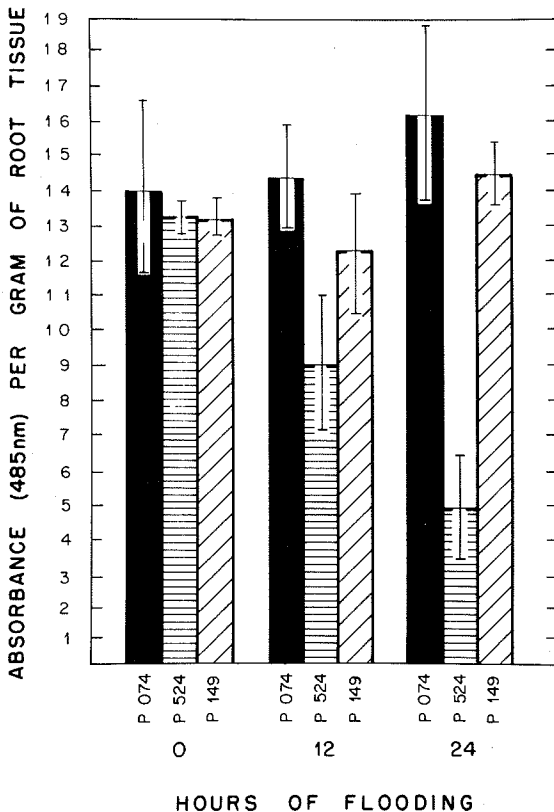


Fig. 1. The effect of flooding for 12 or 24 h as compared to no flooding (0 h) on the ability of bean seedling root-tip segments from genotypes P074, P524 and P149 to reduce TTC. Increasing ability to reduce TTC indicates increasing flooding tolerance. Error bars indicate the mean  $\pm$  1 standard deviation of 3 samples from each genotype within each time sequence.

TABLE I

Summary of the response of 3 bean genotypes to 4 methods of measuring flooding tolerance

Genotype	Method of measurement			
	TTC	EC	PC	Visual index
P524	Tolerant	Tolerant	Tolerant	Tolerant
P074	Sensitive	Sensitive	Semi-sensitive	Sensitive
P149	Tolerant	Tolerant	Sensitive	Sensitive

The flooding of soil decreases the supply of oxygen to roots, as compared normally-aerated soil, by a factor of  $3 \times 10^6$  (Drew, 1979) and therefore inhibits aerobic respiration of the living tissue. TTC reduction is thought to indicate the status of oxidation-reduction reactions in the cell (Palta et al., 1981). The measure of TTC reduction ability can thus be taken as indicative of the rate of respiration in the tissue. It can be expected that during flooding of plants, the tolerance to waterlogging will be directly related to the extent of TTC reduction ability maintained by the tissue. A 50% loss in TTC reduction ability has been found to be approximately related to the irreversible injury produced by the freezing and heat stress in potato species (Palta et al., 1981). It is possible that 24 h of flooding stress produced irreversible injury in P524 and no significant injury in P074 and P149.

*method.* — In general, the response trend of the genotypes to waterlogging over the 36-h period was similar to that of the TTC experiment, with P524 susceptible and P074 and P149 tolerant, with a significant difference between the 2 groups (Fig. 2). The length of time required for a plant tissue sample to lose 50% of its cell solutes to the surrounding environment is generally understood as the critical point of stress where the tissue sample is unable to recover to a viable, functional condition (Palta et al., 1978). This stage was attained only by P524 after 8 h. P524 was also the only genotype to reach a "non-viable" condition upon prolonged flooding (visual observations). As in the TTC experiment, P074 demonstrated greater tolerance ( $P \leq 0.05$ ) than P149 only between the 12- and 24-h flooding treatments.

Both flooding duration and genotype influenced ( $P \leq 0.005$ ) the amount of cell solutes leached from root segments. Plant-to-plant differences were found ( $P \leq 0.005$ ) within each time and genotype, and there was low variability among the 3 root segments within plants.

*method.* — Xylem water potential provided good differentiation ( $P \leq 0.005$ ) among the 3 genotypes (Fig. 3). As shown by the TTC and EC methods, P074 was more tolerant than P524 or P149. Neither plant age over a period

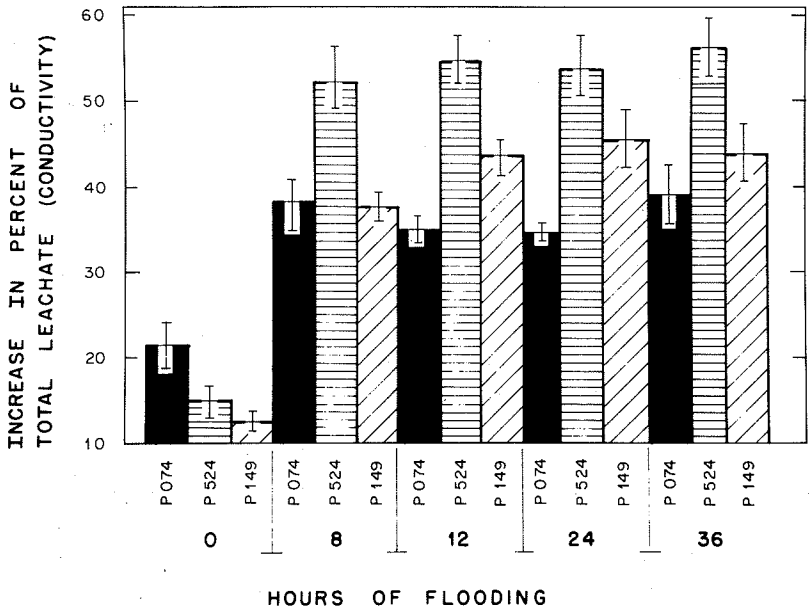


Fig. 2. The effect of flooding upon the percent of total cell solutes leached from seed root segments from bean genotypes P074, P524 and P149. Increasing amounts of contents leached indicates decreasing flooding tolerance. Each bar represents the  $\pm 1$  standard deviation of 9 segments from 3 seedlings.

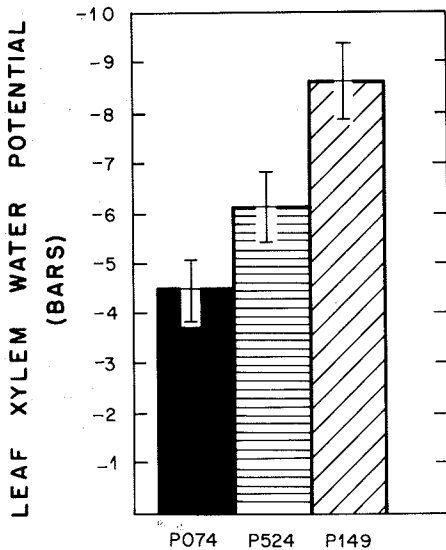


Fig. 3. The effect of 70 h of flooding on the plant xylem water potential of 3 bean types. Decreasing water potentials indicate decreasing flooding tolerance. Bars represent mean  $\pm 1$  standard deviation of 4 replications of 2 plants each with 3 samples per

2 weeks nor leaf samples within plants caused xylem water potential change. These observations indicate that (1) a uniform performance within genotype may be expected when measuring the xylem water potential of plants with the PC under growth-chamber conditions for a period of about 2 weeks, and (2) the sampling of a single leaf may be expected to provide a representative indication of the water status of other parts of the leaf canopy. The degree of relationship between the visual scoring and the xylem water potential across the 3 genotypes after 70 h of flooding was as high as 0.85. This is evidence for the utilization of either method in the determination of tolerance.

*Comparison among methods.* — While P074 and P524 were found to be very tolerant and sensitive, respectively, by all of the 4 methods, the response of P149 was not as consistent (see Table I). P149 was relatively tolerant according to the TTC and EC methods, but sensitive according to the PC and visual scoring methods. This apparent inconsistency may be due to the possibility that each method measures a particular physiological component contributing to whole plant tolerance, which conceivably could differ in its expression or cause, even between cultivars of the same species (Crawford, 1966). Presumably, the visual scoring method might be used to compare the effectiveness of the other methods. The precision and sensitivity of the TTC, EC and PC methods make them practical indicators of the underlying physiological condition of the stressed plant.

For rapid use as screening tools enabling selection of large numbers of genotypes in a breeding program, TTC and EC, as now used, may be too laborious. They would also require more highly trained personnel. The measurement of plant desiccation via the PC might be relatively useful, however. The mobility of the instrument, and the non-destructivity and repeatability of the method for each plant make it attractive to the plant breeder. Although no prior studies regarding its utilization for measuring chlorophyll damage are known, strong relationships have been found between water potential measurements and other methods used to evaluate flooding tolerance, such as transpiration rates (Regehr et al., 1975), the percentage of wilted plants (Rowe and Catlin, 1971), and levels of abscisic acid and ethylene (Wright, 1977). However, our results indicate that visual scoring may also be a satisfactory method. Both the visual and PC methods are non-destructive, but they do require larger, well-developed plants and full environmental control during plant growth and flooding. The TTC and EC methods, on the other hand, are suitable for use with seedlings, significantly reducing the time and space required to grow plants for screening purposes. Nevertheless, it should be remembered that sensitivity to flooding could be different at different plant-growth stages.

*Inheritance study.* — In the inheritance study, differences between P524 and P149 and among the populations generated from them were non-sig-



nificant; hence our analysis of them did not proceed further. There was considerable phenotypic variability within both P074 and P149, based on the visual rating. Only 62% of the P074 plants were rated tolerant, whereas 85% of the P149 plants were rated sensitive. Segregation within the  $F_1$  and back-cross populations appeared to fit a normal distribution. Because sample sizes were small and the genetic constitution of the parents not known, firm conclusions could not be made regarding the inheritance of flooding tolerance (Fig. 4). However, back-cross differences ( $P \leq 0.05$ ) were found between  $(P074 \times P149) \times P074$  and  $P074 \times (P074 \times P149)$ . The back-cross of the  $F_1$  to P074 appeared to transfer a degree of tolerance which exceeded that of the  $F_1$ . The back-cross of the  $F_1$  to P149 appeared to transfer a degree of flooding sensitivity greater than that of the  $F_1$ .

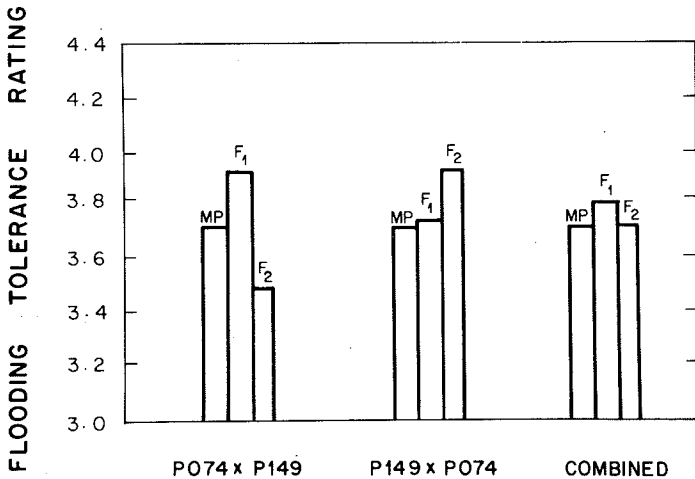


Fig. 4. Mid-parent (MP),  $F_1$  and  $F_2$  mean visual index ratings for flooding tolerance in reciprocal bean crosses. Increasing index value indicates decreasing flooding tolerance as based on a 1-6 scale.

## CONCLUSION

Our study shows that several methods can be used to evaluate the degree of stress imposed on *Phaseolus vulgaris* genotypes differentially sensitive to soil waterlogging. PC and visual scoring methods were equally effective. EC and EC were also useful, but were more time-consuming and gave more variable results. However, the latter 2 methods require only 7-day old seedlings, thus permitting the use of larger samples and more efficient use of space.

## REFERENCES

- Boyer, J.S., 1967. Leaf water potentials measured with a pressure chamber. *Plant Physiology* 42: 133-137.

- ne, P., Marshall, D.R. and Munday, J., 1976. The response of lupins to waterlogging. *Aust. J. Exp. Agric. Anim. Husbandry*, 16: 549-555.
- ro Internacional de Agricultura Tropical, 1977. 1977 Annual Report. Cali-Valle, Colombia, South America, pp. B37-B38.
- ts, M.P. and Philipson, J.J., 1978. The tolerance of tree roots to waterlogging. II. adaptation of Sitka spruce and Lodgepole pine to waterlogged soil. *New Phytol.*, 80: 71-77.
- ford, R.M.M., 1966. The control of anaerobic respiration as a determining factor in the distribution of the genus *Senecio*. *J. Ecol.*, 54: 403-413.
- ford, R.M.M., 1967. Alcohol dehydrogenase activity in relation to flooding tolerance in roots. *J. Exp. Bot.*, 18: 458-464.
- ford, R.M.M. and McManmon, M., 1968. Inductive responses of alcohol and malic dehydrogenases in relation to flooding tolerance in roots. *J. Exp. Bot.*, 19: 435-441.
- er, S.T., Tottingham, W.E. and Graber, L.F., 1932. Investigation of the hardiness of plants by measurement of electrical conductivity. *Plant Physiol.*, 7: 63-78.
- y, M.C., 1979. Plant responses to anaerobic conditions in soil and solution culture. *Curr. Adv. Plant Sci.*, 36: 1-14.
- eva, G.M., 1962. Excretion by plant roots during brief periods of anaerobiosis. *Sov. Plant. Physiol.*, 8: 549-552.
- , F.W. and Zorin, M., 1977. Heritability studies related to waterlogging tolerance in selection of creeping-rooted lucerne. *Aust. CSIRO Div. Plant Ind. Field Stn. Rec.*, 16: 25-31.
- ock, L.D. and Law, A.G., 1968. Relationship of seedling vigor to respiration and tetrazolium chloride reduction by germinating wheat seeds. *Agron. J.*, 60: 286-288.
- ner, P.J., 1951. Causes of injury to plants resulting from flooding of the soil. *Plant Physiol.*, 26: 722-736.
- n, R. and Jerchel, D., 1941. Über invertseifen. VIII. Reduktion von tetrazoliumsalsen durch Bakterien, gärend Hefe und keimende Same. *Ber. Dtsch. Chem. Ges.*, 74: 49-952.
- H. and Palta, J.P., 1978. Frost hardening and freezing stress in tuber-bearing *Solanum* species. In: P.H. Li and A. Sakai (Editors), *Recent Advances in Plant Cold Hardiness and Freezing Stress: Mechanisms and Crop Implications*. Academic Press, New York, pp. 49-71.
- hall, T. and Millington, A.J., 1967. Flooding tolerance of some western Australian pasture legumes. *Aust. J. Exp. Agric. Anim. Husbandry*, 7: 367-371.
- , J.P. and Li, P.H., 1980. Alterations in membrane transport properties by freezing injury in herbaceous plants: Evidence against rupture theory. *Physiol. Plant.*, 50: 159-175.
- , J.P., Levitt, J. and Stadelmann, E.J., 1977. Freezing injury in onion bulb cells. I. Evaluation of the conductivity method and analysis of ion and sugar efflux from injured cells. *Plant Physiol.*, 60: 393-397.
- , J.P., Levitt, J. and Stadelmann, E.J., 1978. Plant viability assay. *Cryobiology*, 15: 19-255.
- , J.P., Chen, H.H. and Li, P.H., 1981. Relationship between heat and frost resistance of several potato species: Effect of cold adaptation on heat resistance. *Bot. Gaz.*, 102: 311-315.
- hr, D.L., Bazzaz, F.A. and Boggess, W.R., 1975. Photosynthesis, transpiration and leaf conductance of *Populus deltoides* in relation to flooding and drought. *Photosynthetica*, 9: 52-61.
- rs, V.E., 1974. The response of lucerne cultivars to levels of waterlogging. *Aust. J. Exp. Agric. Anim. Husbandry*, 14: 520-525.
- e, R.N. and Catlin, P.B., 1971. Differential sensitivity to waterlogging and cyanogenesis by peach, apricot and plum roots. *J. Am. Soc. Hortic. Sci.*, 96: 305-308.

- Scholander, P.F., Hammel, H.T., Hemmingsen, E.A. and Bradstreet, E.D., 1964. Hydraulic pressure and osmotic potential in leaves of mangroves and some other plants. *Proc. Natl. Acad. Sci.*, 52: 119-125.
- Steponkus, P.L. and Lanphear, F.O., 1967. Refinement of the triphenyl tetrazolium chloride method of determining cold injury. *Plant Physiol.*, 42: 1423-1426.
- Torres, A.M., Dieddenhofen, V. and Johnstone, I.M., 1977. The early allele of alcohol dehydrogenase in sunflower populations. *J. Hered.*, 68: 11-16.
- Towill, L.E. and Mazur, P., 1975. Studies on the reduction of 2,3,5-triphenyltetrazolium chloride as a viability assay for plant tissue cultures. *Can. J. Bot.*, 53: 1097-1102.
- Wright, S.T.C., 1977. The relationship between leaf water potential and the level of abscisic acid and ethylene in excised wheat leaves. *Planta*, 134: 183-189.
- Yoshida, S. and Todano, T., 1978. Adaptation of plants to submerged conditions. In G.A. Gung (Editor), *Crop Tolerance to Suboptimal Land Conditions*. Am. Soc. Agron. Special Pub. No. 32, pp. 233-256.

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